# mRNA for chosen pro- and anti-inflammatory cytokines in T-lymphocytes in paediatric leukemias and lymphomas - a preliminary report

Łuczyński W<sup>1</sup>, Kovalchuk O<sup>2</sup>, Krawczuk-Rybak M<sup>1</sup>, Malinowska I<sup>3</sup>, Mitura-Lesiuk M<sup>4</sup>, Matysiak M<sup>3</sup>, Kowalczyk J<sup>4</sup>, Chyczewski L<sup>2</sup>

<sup>1</sup>Department of Paediatric Oncology, <sup>2</sup>Department of Clinical Molecular Biology, Medical University of Białystok; <sup>3</sup>Department of Paediatrics, Haematology and Oncology, Medical University of Warsaw, <sup>4</sup>Department of Paediatric Haematology and Oncology, Medical University of Lublin

## Abstract

We assessed mRNA for chosen pro- and anti-inflammatory cytokines in T-lymphocytes of peripheral blood in paediatric patients with leukemias and lymphomas. Levels of four different cytokine mRNAs (IFN-γ, IL-10, IL-4, TGF-β) were determined by the real-time PCR technique. In the whole examined group, at the time of diagnosis, we noted lower amounts of mRNA for TGF- $\beta_1$ , comparing to respective values in the control patients. In the ALL group, we observed the following: 1) at the time of diagnosis: lower amounts of mRNA for IL-4 and for TGF- $\beta_1$ , comparing to respective values in the control group; 2) lower amounts of mRNA for IL-10 after remission induction, comparing to the time of diagnosis. In our opinion, "immunedysregulation" in lymphoproliferative diseases in children is not caused by IFN-y deficiency. The deficit of antiinflammatory cytokines, i.e., IL-4, TGF-B, with higher amounts of IL-10, suggests their role in cancer development.

Key words: leukaemia, lymphoma, children, mRNA, cytokines, T-lymphocytes

## Introduction

Haematological malignancies represent a unique group of proliferative diseases, in which neoplastic cells arise from the immune system and thus, they exert a potential immunomodulatory effect. Leukaemic blasts, like normal lymphocytes, are produced in bone marrow and these two types of cells may inter-

ADDRESS FOR CORRESPONDENCE: Włodzimierz Łuczyński Department of Paediatric Oncology Medical University of Białystok Waszyngtona 17, 15-274 Białystok, Poland e-mail: vlodek@amb.edu.pl act. It is likely that neoplastic cells produce substances, which impair the immune response, thus promoting cancer expansion. The main role in antitumour immunity is played by T-lymphocytes. According to produced cytokines, T-cells can be differentiated into Th<sub>1</sub>/Tc<sub>1</sub> subsets (producing IL-2 and IFN- $\gamma$ ), Th<sub>2</sub>/Tc<sub>2</sub> (IL-4, IL-5, IL-10) and Th<sub>3</sub>/Tr<sub>1</sub> (TGF-β, IL-10). Most of the authors report a Th2 polarization profile in haematological malignancies and normalization of the Th1/Th2 balance in remission [1]. A type 1-to-type 2 cytokine shift may be responsible for reduced antitumor immunity and is correlated with disease progression. We previously found Th2 (the percentage of CD4+IL-4+ cells - flow cytometry) predominance at the time of diagnosis of acute lymphoblastic leukaemia (ALL) [2]. In the present, study we assessed mRNA for chosen pro- and anti-inflammatory cytokines at the time of diagnosis of most common lymphoproliferative diseases in children.

### Material and methods

Twenty (20) patients with ALL (n=13), Hodgkin lymphoma (n=4) and non-Hodgkin lymphoma (n=3) were prospectively enrolled into this study. Peripheral blood (1 ml) was taken at the time of diagnosis and in ALL - after prednisone prophase (day  $8^{th}$ ), during (day  $15^{th}$ ) and after remission induction (day  $33^{rd}$ ). The control group included 30 children from the Department of Paediatric Surgery, subjected to minor surgical operations.

CD4<sup>+</sup> and CD8<sup>+</sup> cells were isolated from whole peripheral blood by immunomagnetic bead separation (Dynabeads, Dynal Biotech, Oslo, Norway). mRNA was isolated from sorted lymphocyte subpopulations, using Dynabeads mRNA DIRECT Micro Kit (DYNAL) according to the producer's instructions. First strand cDNA was synthesized, using random hexamers as primer and a High Capacity cDNA Archive Kit by AppliedBiosystems. Levels of four different cytokines mRNAs (IFN- $\gamma$ , IL-10, IL-4, TGF- $\beta$ ) were determined by the real-time PCR technique with the TaqMan chemistry, using ready-to-use

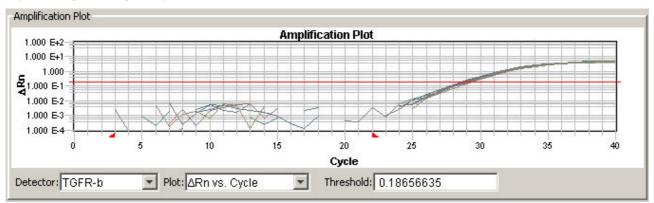


Figure 1. Amplification plot (log view) of TGF-b mRNA

Assays-on-Demand Gene Expression Products by AppliedBiosystems, which contain target-specific primers and probe, and a TaqMan Universal Master Mix, containing AmpErase uracil-N-glycosylase (UNG) to prevent re-amplification of carryover PCR products. PCR amplification and fluorescence data collection were performed with the ABI PRISM 7900HT Sequence Detection System (AppliedBiosystems). In order to normalize the amount of expressed cytokine mRNAs, the internal housekeeping gene GAPDH was used and each complementary DNA (cDNA) product was tested in triplex for each of the four cytokines mRNA and GAPDH mRNA. For the calculation of our data we used the Comparative Ct method for relative quantification (AACt method), which describes the change in expression of the target gene in a test sample relative to a calibrator sample and provides an accurate comparison between the initial levels of template in each sample. We used Total Raji RNA by AppliedBiosystems, as a calibrator sample, which was processed in the same way as the test samples. The obtained data were analysed with the Sequence Detector System (SDS) software, version 2.1 (AppliedBiosystems).

## Results

In the examined groups and in the control group, we found the following: a) in CD4<sup>+</sup> cells: positive correlations between mRNA for IFN-γ and: IL-4 (r=0.83, p<0.0001), TGF-β<sub>1</sub> (0.91, p<0.0001); and between IL-4 and TGF-β<sub>1</sub> (r=0.86,p<0.0001); b) in CD8<sup>+</sup> cells: a negative correlation between mRNA for IFN- $\gamma$  and IL-10 (r=-0.44, p=0.0007); a positive correlation between IFN-γ and IL-4 (r=0.43, p=0.01), IFN-γ and TGF-β<sub>1</sub> (r=0.81, p<0.0001); also a positive correlation between IL-4 and TGF- $\beta_1$  (r=0.7, p<0.0001). In the whole examined group, at the time of diagnosis, we noted lower amounts of mRNA for TGF- $\beta_1$  in CD4<sup>+</sup> cells (the difference not statistically significant) and in CD8<sup>+</sup> cells (p=0.01) comparing to the control patients; the amounts of other cytokines (IFN-y, IL-4, IL-10) did not differ between those in the whole examined group and those in the control patients. In the ALL group, we observed: a) at the time of diagnosis: lower amounts of mRNA for IL-4 (both in CD4+ and CD8<sup>+</sup> cells, p=0.02 and p=0.04) and for TGF- $\beta_1$  (also in both subpopulations p=0.03 and p=0.01), comparing to respective values in the control group; b) lower amounts of mRNA for IL-10 after remission induction, comparing to the time of diagnosis

(p=0.05). An example of amplification plot for TGF- $\beta$  is presented on Figure 1.

## Discussion

The only report, concerning Th<sub>1</sub>/Th<sub>2</sub> balance in children with ALL, was performed by Zhang et al. [3]. The authors found lower Th<sub>1</sub>/Th<sub>2</sub> and Tc<sub>1</sub>/Tc<sub>2</sub> ratios at the time of diagnosis and recovery after achieving complete remission. Regarding IFN-y, we did not find any difference between the examined group and the control. This confirms our previous results, obtained with flow cytometry, and the results obtained by Kiani et al. in CML patients [2, 4]. Podhorecka et al. found a higher Thi percentage in patients with B-cell chronic lymphocytic leukaemia (B-CLL) than in controls and the Th<sub>2</sub>/Tc<sub>2</sub> shift during disease progression [5]. The amounts of mRNA for IL-4 were lower in ALL patients at the time of diagnosis. Similar results, concerning IL-4, were obtained by Kiełbiński et al. in acute leukaemias in adults [6]. In contrast to our results, Kamińska et al. observed, comparable with the control levels of IL-4 in supernatants from the whole blood cell cultures of adult patients with ALL [7]. Mori et al. reported polarization to Th<sub>2</sub> in untreated B-cell diffuse large cell lymphoma patients and to Th<sub>1</sub> in complete remission [1]. We noted a tendency (not significant) to higher amounts of mRNA for IL-10 at the time of diagnosis of leukemias and lymphomas. IL-10 production in patients with cancer was higher than that in healthy controls, so it is possible that tumour cells secrete cytokines, shifting T-cells to produce more IL-10 [8]. Kebelmann-Betzing et al. suggest that the secretion of IL-10 (from leukaemic blasts) and the lack of down-regulation of adhesion and costimulatory molecules determine the mechanism of escape from immune surveillance in relapsed ALL [9]. In the whole examined group and, especially in ALL patients, we observed lower amounts of mRNA for TGF-β, comparing to the values in the control patients. Tatsumi et al. found Th2, but not Th<sub>3</sub>, type response against renal cell carcinoma or melanoma [10]. In our opinion "immunedysregulation" in lymphoproliferative diseases in children is not caused by IFN-γ deficiency. The deficit of anti-inflammatory cytokines, i.e., IL-4, TGF-B and the higher amounts of IL-10, revealed in that group of children, suggests their role in cancer development, but it should be confirmed in a greater cohort of patients. The very strong correlations, observed among all the assessed substances, suggest their interdependence in cytokine network in the human organism. Determination of the factors, that affect Th and Tc profiles, may lead to a better understanding of the immunological status and development of immunotherapy.

The study supported by The State Committee for Scientific Research (project number P05E08025)

#### References

1. Mori T, Takada R, Watanabe R, Okamoto S, Ikeda Y. T-helper (Th)<sub>1</sub>/Th<sub>2</sub> imbalance in patients with previously untreated B-cell diffuse large cell lymphoma. Cancer Immunol Immunother, 2001; 50: 566-8.

2. Łuczyński W, Stasiak-Barmuta A, Krawczuk-Rybak M, Malinowska I, Matysiak M, Mitura-Lesiuk M, Kowalczyk J, Jeromin A. Równowaga limfocytów Th<sub>1</sub>/Th<sub>2</sub> w ostrej białaczce limfoblastycznej u dzieci. Przeg Lek, 2004; in press.

3. Zhang X-L, Komada Y, Chipeta J, Li Q-S, Inaba H, Azuma E, Yamamoto H, Sakurai M. Intracellular cytokine profile of T cells from children with acute lymphoblastic leukemia. Cancer Immunol Immunother, 2000; 49: 165-72.

4. Kiani A, Habermann I, Schake K, Neubauer A, Rogge L, Ehninger G. Normal intrinsic Th<sub>1</sub>/Th<sub>2</sub> balance in patients with chronic phase chronic myeloid leukemia not treated with interferon-alpha or imatinib. Haematologica, 2003; 88: 754-61.

5. Podhorecka M, Dmoszyńska A, Roliński J, Wasik E. T type 1/type 2 subsets balance in B-cell chronic lymphocytic leukemia - the three-color flow cytometry analysis. Leuk Res, 2002; 26: 657-60.

6. Kiełbiński M, Podolak-Dawidziak M, Prajs I, Haus O, Jaźwiec B, Kuliczkowski K. Limfocyty Th<sub>1</sub> i Th<sub>2</sub> w nadzorze odporności w ostrych białaczkach. Acta Hematol Pol, 2003; 34: 360.

7. Kamińska T, Hus I, Dmoszyńska A, Kandefer-Szerszeń M. Cytokine production in whole blood cell cultures of patients with B-lineage acute lymphoblastic leukemia. The influence of granulocyte-macrophage colony-stimulating factor. Arch Imm Ther Exp, 2001; 49:71-7.

8. Kallio R, Surcel H-M, Syrjala H. Peripheral mononuclear cell IL-10 and IL-12 production is not impaired in patients with advanced cancer and severe infection. Cytokine, 2002; 20: 210-4.

9. Kebelmann-Betzing C, Korner G, Badiali L, Buchwald D, Moricke A, Korte A, Kochling J. Characterization of cytokine, growth factor receptor, costimulatory and adhesion molecule expression patterns of bone marrow blasts in relapsed childhood B cell precursor ALL. Cytokine, 2001; 13: 39-50.

10. Tatsumi T, Kierstaed LS, Ranieri E, Gesualdo L, Schena FP, Finke JH, Bukowski RM, Mueller-Brghaus J, Kirkwood JM, Kwok WW. Disease-associated bias in T helper type 1 (Th<sub>1</sub>)/Th<sub>2</sub> CD4<sup>+</sup> T cell responses against MAGE-6 in HLA-DRB1\*0401<sup>+</sup> patients with renal cell carcinoma or melanoma. J Exp Med, 2002; 195: 619-29.