

# Effect of hydralazine on CD3- $\zeta$ chain expression in Jurkat T cells

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## Abstract

**Purpose:** Deficient CD3- $\zeta$  chain expression in T cells of patients with idiopathic SLE is associated with T cell receptor/CD3 complex (TCR/CD3)-mediated signaling defect. Hydralazine (HYD) inhibits expression of DNA methyltransferase 1 (DNMT1) and may cause a lupus-like disease in man.

**Material and methods:** To explain the HYD effect on intracellular level of CD3- $\zeta$  chain in Jurkat T leukemia cells clone E6-1, we employed the flow cytometric analysis.

**Results:** We observed a dose-dependent increase in cellular content of CD3- $\zeta$  chain in Jurkat T cells treated with HYD. Our results suggest that HYD may result in T cells dysfunction different from this observed in idiopathic SLE T cells.

**Conclusions:** This difference may partially explain distinct disease course in patients with HYD induced and idiopathic SLE.

**Key words:** hydralazine, T lymphocytes, cell signalling, DNA methylation.

## Introduction

Systemic lupus erythematosus (SLE) is autoimmune disease characterised by abundant production of autoantibodies. Defect in CD4<sup>+</sup> T lymphocytes signaling can be responsible for improper immune response development in patients with SLE [1].

The T cells stimulation is initiated by binding TCR/CD3 with an antigen coupled to the major histocompatibility complex [2]. The  $\zeta$  chain is component of CD3 complex and plays a major role in intracellular signaling transduction, which activate second messenger and transcription factors [2-4]. T cells stimulation induces cytokines production, increases proliferation and augmentation of effector function of T cells [2].

The defect in T cells signalling can be responsible for improper immune response development in patients with SLE [5].

It has been reported that low methylation of CpG residues in the regulatory sequences of DNA and high level of histone acetylation correlate with transcriptional activity of numerous genes [6-8]. During DNA replication, the CpG pairs of the newly biosynthesised DNA strand are methylated by DNA methyltransferase 1 (DNMT1) [6]. Expression of DNMT1 is partially regulated by extracellular signal regulated kinase pathway (ERK), and activity of this pathway is decreased in T cells from SLE patients [9]. Hydralazine (HYD) is a substance, which is able to induce a lupus-like syndrome in man. HYD inhibits ERK pathway resulting in decrease of DNMT1 expression and DNA hypomethylation [10].

Using the flowcytometric analysis, we evaluated the effect of HYD on CD3- $\zeta$  chain content in Jurkat T leukemia cells.

## Material and methods

### Reagents and Antibodies

HYD and digitonin were obtained from Sigma Chemical Co. (St. Louis, MO). (PE)-conjugated anti-CD3- $\zeta$  (6B10.2) mouse monoclonal antibody (MmAb) was purchased from Santa Cruz Biotechnology, Inc. USA.

### Cell culture and HYD treatment

Jurkat T leukemia CD4<sup>+</sup> cells clone E6-1 were obtained from the American Type Culture Collection (Rockville, MD) and maintained in RPMI1640 (GibcoBRL, Grand Island, NY)

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medium containing 10% heat-inactivated foetal bovine serum (FCS), 2 mM glutamine, 100  $\mu\text{g}/\text{ml}$  streptomycin, and 100 U/ml penicillin. Jurkat T leukemia cells were suspended at a concentration  $0.5 \times 10^6$  cells/ml in culture medium and grown for 48 h without, or in the presence of HYD in concentration of 0.5, 5.0 and 50.0  $\mu\text{M}$ .

#### Flow cytometric analysis

After incubation, the cells were harvested and washed three times in phosphate buffered saline (PBS) supplemented with 2% FCS and 1% sodium azide (PBS/FCS).

The cells were permeabilized with digitonin 10  $\mu\text{g}/\text{ml}$ , fixed with 0.25% paraformaldehyde and washed three times with PBS/FCS. The cells were then stained with PE-conjugated anti CD3- $\zeta$  MmAb, washed three times with PBS/FCS and immediately analysed on FACSCanto Flow Cytometer (Becton-Dickinson, San Jose, CA). The increase of CD3- $\zeta$  chain cellular content were calculated according to  $(\text{MFx}-\text{MFO})/(\text{MFC}-\text{MFO}) \times 100$  formula. MF is the mean fluorescence intensity of cells, which were grown in the presence (MFx) or absence (MFC) of HYD and then stained with PE-conjugated anti CD3- $\zeta$  MmAb. Control represent fluorescence intensity of cell stained with an appropriate isotope antibody (MFO).

## Results and discussion

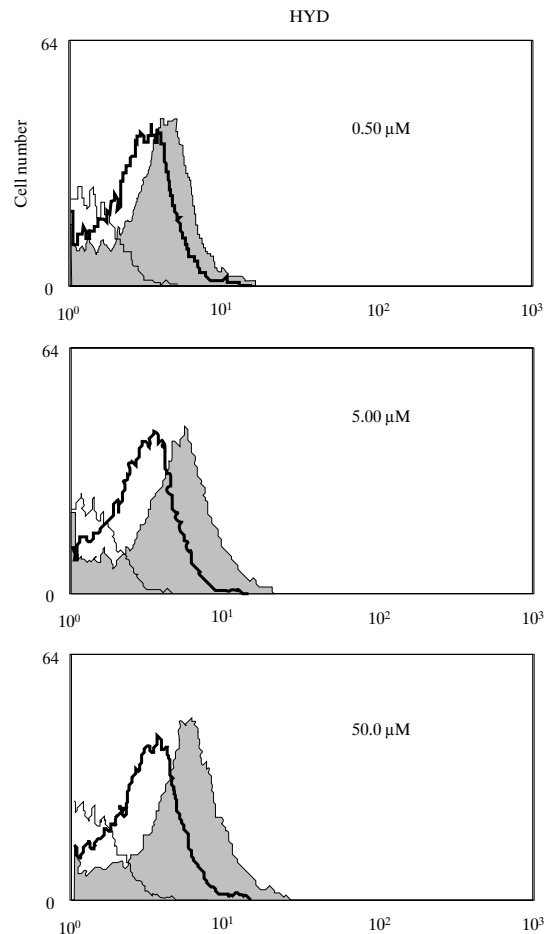
In order to explain the HYD effect on intracellular level of CD3- $\zeta$  chain in Jurkat T leukemia cells, we employed the flow cytometric analysis. We observed that the HYD increased intracellular contents of CD3- $\zeta$  chain in Jurkat T cells in dose dependent manner. We have shown that the percentage increase of CD3- $\zeta$  chain formation achieves  $17 \pm 3\%$ ,  $70 \pm 13\%$  and  $238 \pm 40\%$  in the presence of HYD concentration of 0.5, 5.0 and 50.0  $\mu\text{M}$ , respectively (Fig. 1).

HYD is used to reduce the blood pressure but may also cause a lupus-like disease in man [10]. The etiology of idiopathic and HYD induced SLE is still enigmatic. Patients with idiopathic and HYD induced SLE exhibit different disease course. However, the T cells from idiopathic and drug induced SLE patients may display certain common defects at molecular level [11-13]. HYD induces biosynthesis of leukocyte function associated-1 (LFA-1) in T cells [10]. The biosynthesis elevation of LFA-1 is also observed in T cells from patients with idiopathic SLE [7]. T cells from this patients also exhibit TCR/CD3-mediated signalling aberrations, which are associated with cellular decrease of CD3- $\zeta$  transcript and protein contents.

The HYD inhibits DNMT1 expression and cause hypomethylation of regulatory DNA sequences that makes DNA template available for transcription [10,14]. The HYD causes hypomethylation of regulatory sequences of LFA-1 gene in T cells, which was also observed in the same region of DNA of SLE T cells [15]. However, we observed that HYD increased CD3- $\zeta$  protein but not transcript content (results not shown) in Jurkat leukemia CD4<sup>+</sup> T cells (Fig. 1). The reason for such an HYD effect on CD3- $\zeta$  chain translation is currently not known. We presume that HYD may effect on factors involved in positive control of translation or posttranslational modification of CD3- $\zeta$  chain.

**Figure 1.** The representative picture of flow cytometric analysis of intracellular contents of CD3- $\zeta$  chain in Jurkat T leukemia cells incubated in the presence of HYD.

Jurkat T leukemia cells clone E6-1 were suspended at a concentration of  $0.5 \times 10^6$  cells/ml in culture medium and were grown for 48 h either without or in the presence of HYD (0.5, 5, 50  $\mu\text{M}$ ). After incubation the cells were harvested, washed with PBS/FCS, permeabilized with digitonin 10  $\mu\text{g}/\text{ml}$  and fixed with 0.25% paraformaldehyde. The cells were then stained with PE-conjugated anti CD3- $\zeta$  MmAb and immediately analyzed on FACSCanto Flow Cytometer (Becton-Dickinson, San Jose, CA). (—) and (---) represent expression of CD3- $\zeta$  chain in cells incubated without or with HYD. Shadow lines represents the cells stained with an appropriate isotype control antibody



Increase of CD3- $\zeta$  chain content in T cells may partially explain different disease course in patients with HYD induced and idiopathic SLE. HYD may also change other elements of TCR signaling pathway resulting in dysfunction of T cells.

The further investigation of HYD effect on expression of other signaling molecules may provide valuable information about etiology of T cells dysfunction in patients with HYD induced SLE.

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