

The impact of genetic factors on response to anaesthetics

Mikstacki A¹, Skrzypczak-Zielinska M², Tamowicz B¹, Zakerska-Banaszak O^{3,4}, Szalata M^{2,3},
Slomski R^{2,3,*}

1 Department of Anaesthesiology and Intensive Therapy, Regional Hospital, Poznan, Poland
2 Institute of Human Genetics, Polish Academy of Sciences in Poznan, Poznan, Poland
3 Department of Biochemistry and Biotechnology, University of Life Sciences, Poznan, Poland
4 The NanoBioMedical Centre, Adam Mickiewicz University, Poznan, Poland

* CORRESPONDING AUTHOR:

Institute of Human Genetics,
Polish Academy of Sciences,
Strzeszynska 32,
60-479 Poznan, Poland
Tel.: +48 61 657 92 10
Fax: +48 61 848 72 11
E-mail: slomski@up.poznan.pl (Ryszard Slomski)

Received 05.06.2012
Accepted 23.11.2012
Advances in Medical Sciences
Vol. 58(1) 2013 · pp 9-14
DOI: 10.2478/v10039-012-0065-z
© Medical University of Białystok, Poland

ABSTRACT

In recent years, exceptional progress has been observed in pharmacogenetics, i.e. investigations of inherited conditioning of the organism's response to drugs or xenobiotics. On the other hand, modern molecular biology techniques have been implemented, making it possible to perform studies determining the involvement of genetic factors in differing responses to agents employed in general anaesthesia. Unexpected and incorrect response of the organism to the administration of specific anaesthetics is most commonly associated with a genetic defect of the metabolic pathway of a given agent or its receptor.

The majority of agents used in anaesthesia are metabolised in the liver by the cytochrome P450 superfamily enzymes (CYPs) and phase II drug-metabolising enzymes: glutathione S-transferases (GSTs), sulphotransferases (SULTs), UDP-glucuronosyltransferases (UGTs) and NAD(P)H:quinone oxidoreductase (NQO1). Propofol is presently widely used for gastrointestinal (GI) and several other procedures. Among genes associated with metabolism of the most commonly applied anaesthetics such as propofol and sevoflurane, the following ones can be mentioned: *CYP2E1*, *CYP2B6*, *CYP2C9*, *GSTP1*, *UGT1A9*, *SULT1A1* and *NQO1*. Moreover, the basic mechanism of propofol action involves its interaction with an ionotropic receptor GABAA inhibiting transfer of nerve impulses. Molecular studies have shown that polymorphic changes in *GABRG2* receptor gene turn out to be important in the propofol anaesthesia.

Planning of optimal anaesthesia can be considerably assisted by the determination of genetic factors of prognostic value taking advantage of genotyping and making it possible to select anaesthetics and reduce risk of side effects as well as undesirable actions.

Key words: anaesthesia, pharmacogenetics, propofol, sevoflurane, genes

INTRODUCTION

First discoveries combining pharmacology, anaesthesiology and genetics appeared shortly after 1959 when a German researcher F. Vogel proposed and defined the concept of pharmacogenetics as a science of genetically conditioned responses to drugs or xenobiotics. At the beginning of 1960s,

Simpson and Kalow [1] in their studies on a local anaesthetic – procaine, and on a muscle relaxing drug – succinylcholine, discovered polymorphism of hydrolysis associated with plasma pseudocholinesterase which fails to guarantee appropriately fast metabolism of the agent. In patients suffering from cholinesterase enzymopathy, an appropriate dose of succinylcholine can lead to undesirable side effects due to excessively long biotransformation [2].

A consecutive breakthrough in the investigations regarding genetic diversity in anaesthetics metabolism was the discovery of the role of drugs and xenobiotics oxidation processes. Oxidised substances become inactive and can be eliminated from the body in the form of hydrophilic products [3]. Detailed identification of metabolic pathways made it possible to determine genes coding enzymes involved in drug metabolism.

The above discoveries were accompanied by the development of nucleic acid analysis methods with its peak at the turn of the 21st century crowned by sequencing of the human genome in 2003 and characterization of over 3.1 million human single nucleotide polymorphisms (SNP) [4-5]. Automation of genetic material amplification techniques, special equipment for genotype determination and mass reading of nucleic acid sequences (microarrays, new generation sequencers), as well as, bioinformatics advancement, all show new possibilities for the application of research results in anaesthesiology. They are based on the determination of alleles closely associated with poor, intermediate, ultra rapid or efficient metabolism of agents currently applied in general anaesthesia. Both, this knowledge and the advancement, aim at the development of a genetic diagnostic test, which will help to individualize anaesthesia for each patient.

REVIEW

General anaesthesia

An ideal anaesthetic agent should be characterised by both anaesthetic and analgesic actions, without any adverse effect on the respiratory and circulatory system, lack of irritating influence on the skin and mucous membrane, wide therapeutic range, lack of transformation to toxic metabolites and easy management of the course of anaesthesia. None of the currently available anaesthetic agents meet the above-mentioned requirements completely.

Therefore, apart from exclusively inhaled or intravenous anaesthesia TIVA (total intravenous anaesthesia), mixed anaesthesia is also frequently employed, combining several agents according to the type of surgery and associated requirements, but also depending on the health condition and age of the patient.

From among inhaled anaesthetic agents, halogen ethers such as: isoflurane, sevoflurane and desflurane introduced in 1990s are applied most commonly [6].

Currently employed halogen ethers contain fluorine (e.g. sevoflurane and desflurane) or chlorine atoms (in the case of isoflurane molecule) in their structure. Chemical composition of halogen ethers does not differ from the old generation agents, except that they possess a higher number of halogen atoms and greater molecular weight.

Sevoflurane (1,1,1,3,3,3-hexa-fluoro-2-(fluoromethoxy)-propane, C₄H₃F₇O) is believed to be an agent of the most

advantageous anaesthetic properties in comparison with the remaining halogen ethers and, therefore, it is applied most frequently in practice [6]. It is absorbed by air vesicles and is characterised by rapid induction time. Anaesthetic action, depending on the concentration of the inhaled sevoflurane, occurs after 1-2 minutes and it also shows a weak analgetic action.

In the case of intravenous general anaesthesia, one of the most commonly applied agents is propofol (2,6-di-isopropylphenol, C₁₂H₁₈O) [7]. It was introduced into the clinical practice in 1970s. An important advantage of these anaesthetics is its short time of introduction into deep anaesthesia (about 30 to 50 seconds) with a possibility of quick waking of the patient (about 4 to 6 minutes).

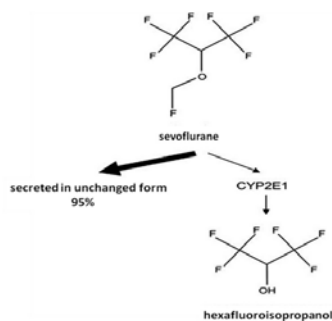
However, in general anaesthesia, the application of potentially most advantageous anaesthetic agents at recommended doses, sometimes turns out to be impossible because side effects such as: bradycardia, hypotension, motoric disorders are observed or even potentially fatal Propofol Infusion Syndrome (PRIS) may occur [8, 9]. The patient's clinical condition exerts an essential influence on the occurrence of undesirable side effects, in particular, serious concomitant diseases may interfere with the course of anaesthesia. However, at present, special attention is focused on the individual metabolic variability that is more and more often reported in literature. It is maintained, on the basis of current research, that the metabolism of the applied substances depends on genetic polymorphisms of enzymes taking part in the biotransformation of the anaesthetic agent or mediating its action, such as receptor proteins [10, 11].

Metabolism of anaesthetics and genetic variants

Sevoflurane. Majority of drugs (about 70-80%) is metabolised in the liver by enzymes from the group of cytochromes P450 (CYPs) and this refers also to anaesthetic agents. CYP genes mutations can cause cancellation, decrease, change or increase of enzyme activity [10] and they belong to the first phase of response. In the case of sevoflurane and isoflurane, enzyme coded by *CYP2E1* gene (cytochrome P450, family 2, subfamily E, polypeptide 1, MIM 124040) take part in their metabolism. Under the influence of *CYP2E1*, approximately 5% of sevoflurane undergoes biotransformation to hexafluoroisopropanol and fluorides [8, 12]. It is suspected that the fluorides may exhibit nephrotoxic action. A toxic effect is also caused by vinyl ether fluoromethyl-2,2-difluoro-1-[trifluoromethyl], a product of sevoflurane degradation following interaction with carbon dioxide absorbents [13]. The remaining 95% of sevoflurane is secreted from the organism in unchanged form (Figure 1).

Thirteen variants of *CYP2E1* gene (Human Cytochrome P450 Allele Nomenclature Committee, www.cypalleles.ki.se/cyp2e1.htm) have been described in literature and the ones occurring most frequently include: *CYP2E1**5 (-1293G>C; -1053C>T) leading to enhanced transcription

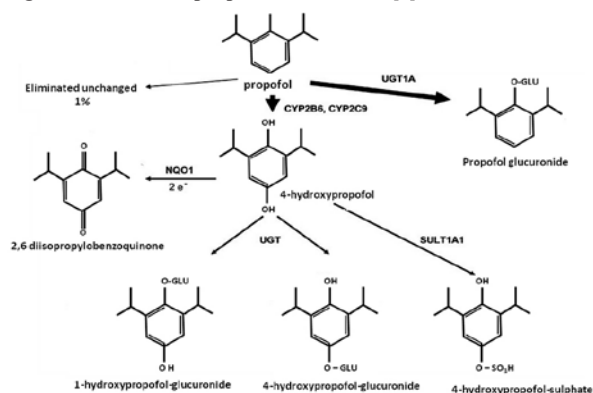
Figure 1. Scheme of sevoflurane metabolism [8].



and *CYP2E1**2 (R144C) reducing enzyme activity [8, 14, 15] (Table 1). Moreover, based on a comprehensive database SuperCYP described by Preissner et al., products of *CYP2A6*, *CYP2B6* and *CYP3A4* genes are also involved in sevoflurane metabolism. [16]. Sevoflurane serves as a substrate for these three cytochrome P450 enzymes [17].

In studies involving the application of inhalatory anaesthetics for general anaesthesia, a close correlation of I105V polymorphism in the glutathione S-transferase gene (*GSTP1*, MIM 134660) with a hepatotoxic effect of these agents was suggested [18]. One of the causes of the toxic action on hepatocytes is the reduction of the liver blood flow. Another mechanism implies biotransformation to toxic metabolites. Measurement of GST enzyme concentration in

Figure 2. Scheme of propofol metabolism [8].



the serum is one of the most sensitive indicators of the liver function and the level of enzyme expression, according to principles of molecular biology, is conditioned by the coding gene genotype, in this case - *GSTP1* [18].

Propofol. Sevoflurane undergoes only slight biotransformation (about 5%) in the organism, whereas propofol is metabolised in the liver in over 90% into a number of products which are secreted with urine. The above biotransformation may run in different ways. Majority of propofol (about 70%) is metabolised into propofol glucuronide, for which UDP-glucuronosyltransferase coded by *UGT1A9* (UDP glucuronosyltransferase 1 family, polypeptide A9, MIM 606434) gene is responsible [19]. An alternative pathway of propofol biotransformation

Table 1. List of most frequent genetic variants associated with sevoflurane and propofol metabolism.

Agent	Gene	Genetic variant	Name of allele	Literature
Sevoflurane/ Isofluran	<i>CYP2E1</i>	R76H	<i>CYP2E1</i> *2	[8, 14, 15]
		-1293G>C; -1053C>T	<i>CYP2E1</i> *5	
Sevoflurane	<i>GSTP1</i>	I105V	---	[18]
	<i>GABRG2</i>	Y444W	---	[28]
	<i>UGT1A9</i>	M33T	<i>UGT1A9</i> *3	[8, 35, 36, 37, 38, 39]
		Y242X	<i>UGT1A9</i> *4	
		D256N	<i>UGT1A9</i> *5	
IVS1+399C/T		---		
Propofol	<i>CYP2B6</i>	K262R (rs2279343)	<i>CYP2B6</i> *4	[8, 39, 40, 41]
		R487C (rs3211371)	<i>CYP2B6</i> *5	
		Q172H (rs3745274)	<i>CYP2B6</i> *9	
		I328T (rs28399299)	<i>CYP2B6</i> *18	
		Q172H + K262R	<i>CYP2B6</i> *6	
		K262R + I328T	<i>CYP2B6</i> *16	
	<i>CYP2C9</i>	R144C I359L	<i>CYP2C9</i> *2 <i>CYP2C9</i> *3	
	<i>SULT1A1</i>	R213H (rs9282861)	<i>SULT1A1</i> *2	[42]
	<i>NQO1</i>	P187S (rs1800566)	<i>NQO1</i> *2	[43]
		IVS4-3C/T	---	

(approximately 29%) is performed by the enzymes coded by *CYP2B6* (MIM 123930) and *CYP2C9* (MIM 601130) genes as well as by *SULT1A* (MIM 171150) and *NQO1* (MIM 125860) genes (Figure 2). So far experiments indicate a relationship between patients' response to propofol in general anaesthesia and polymorphism of these genes (Table 1). In addition, there are suggestions in the literature about other polymorphisms located in the promoter region, which may play an important role in the enzyme activity and propofol biotransformation, for example -118(dT)⁹>10, -275(T>A)/-2152(C>T) in the *UGT1A9* gene [20, 21]. Following the action of *CYP2B6* and *CYP2C9* enzymes, a 4-hydroxypropofol develops and the end-products include: 2,6-diisopropyl-1,4-benzoquinone with the *NQO1* participation, 1- and 4-hydroxypropofol 1-O- β -D-glucuronide with the *UGT* participation as well as 4-hydroxypropofol sulphate as a result of the action of the *SULT1A* enzyme [19, 22].

It is well known that the basic mechanism of propofol action is based on its interaction with an ionotropic receptor $GABA_A$ inhibiting the transfer of nerve impulses between neurons in the central nervous system [23]. The $GABA_A$ (gamma-aminobutyric acid type A) receptor involved in propofol action is a protein of complex structure. The α , β , γ , δ , ϵ , θ , ρ , π subunits have been discovered which, in various combinations, may participate in the composition of the receptor and, hence, determine its sensitivity to $GABA$ [24]. The dominant receptor isoform in the central nervous system consists of $\alpha 1$, $\beta 2$ and $\gamma 2$ subunits. The activity of the receptor is regulated by the binding of a specific ligand – γ -aminobutyric acid (in particular, presence of $\gamma 2S$, $\gamma 2L$, $\beta 2$ and $\alpha 2$ subunits enhances affinity) but it also contains domains recognizing anaesthetics. In the case of propofol, these include mainly $\beta 3$ and $\beta 2$ subunits but also $\beta 1$ [25]. Receptor activation, which results in the hyperpolarisation of the neuron membrane and, hence, prevents the development of an action potential, takes place as a result of the intensification of the influence of the γ -aminobutyric acid on $GABA_A$ or by way of a direct induction caused by the anaesthetic agent [26]. The performed investigations proved that the activation of the receptor as a result of propofol action (3 $\mu\text{g/ml}$) was the strongest after 2 minutes and the hyperpolarisation of the neuron membrane lasted up to 10 seconds.

Genes coding $GABA_A$ receptor subunits are situated in cluster forms on: 5q34, Xq28, 4p12 and 15 chromosomes. So far, 19 genes have been discovered including, among others: *GABRB2* coding subunit $\beta 2$ as well as *GABRA1*, *GABRA3*, *GABRG1*, *GABRG3*, *GABRB3* genes [27]. In literature, polymorphism of selected genes is particularly stressed in the action of propofol but the current knowledge in this area is superficial. Investigations indicate that the course of general anaesthesia is also affected by *GABRG2* (gamma-aminobutyric acid, MIM 137164) gene polymorphism and, in particular, by the change of amino acid tyrosine to tryptophan at codon 444 (Y444W) [28] (Tab. 1). The four

polymorphic variations (358G/T, 20118C/T, 20326C/T and 20502 A/T) in the *GABRE* gene showed no statistically significant correlation with the anaesthesia induction time, but the impact of this gene on propofol anaesthesia cannot be excluded [29].

Symptoms of undesirable action of propofol include: short-term apnoea changing into hyperventilation, muscular tremors, drop of blood pressure as well as vision disturbances and hallucinations [9].

Perspectives of development and application of pharmacogenetics in anaesthesiology

Population genetic variability has already been described many times using as examples various disorders as well as differences in drug metabolism [30]. Such information is extremely useful and frequently makes it possible to narrow the scope of search in molecular-genetic diagnosis. However, when analysing response to treatment, it is necessary to remember not only about differences at the level of populations but also of each organism. This principle also refers to the administration of agents for general anaesthesia where it is important to individualise the use of anaesthetics in order to ensure optimal effect for each anaesthetised patient [31]. A significant support in this field can be provided by precise indication of genetic factors taking part in the metabolism of anaesthetics as well as analgetic agents.

Scientists have already determined genes taking part in metabolism of individual substances which was also shown in this study taking as examples the most frequently administered anaesthetics – sevoflurane and propofol. In addition, appropriate tools have also been developed such as the haplotype map (HapMap) intended, in particular, for investigations in the fields of pharmacogenetics and pharmacogenomics [32]. The above-mentioned achievements, in association with further clinical studies of genetic factors in anaesthesiology may result, in near future, in the development of a diagnostic tool for precise and individualised anaesthetics adjusted to patients' genotypes. In close perspective, the next challenge will be combining a genetically conditioned response to an agent with the impact of environment, i.e. moving research into the field of pharmacoepigenomics [33, 34].

CONCLUSION

In recent years, pharmacogenetics has been the object of intensive study for many branches of medicine. It may be the basis of personalized anaesthesiology in the near future. Determination of genetic factors of prognostic value for a target anaesthetic agent choice and dosing, before the start of surgery, would improve the safety of patients especially with cardiac or renal dysfunction. Anaesthetics biotransformation in the organism as well as genes encoding

proteins of metabolic pathways are known. Scientists have already done research on the impact of single genotypes on the pharmacokinetics and pharmacodynamics of propofol and on a hepatotoxic effect of sevoflurane in the patients under surgical anaesthesia. However, now it is important to carry out further comprehensive clinical studies in different populations, and hopefully, it will be possible to introduce dosing recommendation for anaesthetics in clinical practice based on individual genotype of patients.

ACKNOWLEDGMENTS

We thank the Polish Ministry of Science and Higher Education for support (grant no. N N401 037838).

REFERENCES

1. Simpson NE, Kalow W. The "Silent" Gene For Serum Cholinesterase. *Am J Hum Genet.* 1964 Jun;16:180-8.
2. Kalow W. Pharmacogenetics and Anesthesia. *Anesthesiology.* 1964 May-Jun;25:377-87.
3. Klinger W. Biotransformation of drugs and other xenobiotics during postnatal development. *Pharmacol Ther.* 1982;16(3):377-429.
4. International Human Genome Sequencing Consortium. Finishing the euchromatic sequence of the human genome. *Nature.* 2004 Oct 21;431(7011):931-45.
5. The International HapMap Consortium. A second generation human haplotype map of over 3.1 million SNPs. *Nature.* 2007 Oct 18;449(7164):851-861.
6. Smith I, Nathanson M, White PF. Sevoflurane—a long-awaited volatile anaesthetic. *Br J Anaesth.* 1996 Mar;76(3):435-45.
7. Watson KR, Shah MV. Clinical comparison of 'single agent' anaesthesia with sevoflurane versus target controlled infusion of propofol. *Br J Anaesth.* 2000 Oct;85(4):541-6.
8. Restrepo JG, Garcia-Martín E, Martínez C, Agúndez JA. Polymorphic drug metabolism in anaesthesia. *Curr Drug Metab.* 2009 Mar;10(3):236-46.
9. Fudickar A, Bein B. Propofol infusion syndrome: update of clinical manifestation and pathophysiology. *Minerva Anesthesiol.* 2009 May;75(5):339-44.
10. Ingelman-Sundberg M. Polymorphism of cytochrome P450 and xenobiotic toxicity. *Toxicology.* 2002 Dec 27;181-182:447-52.
11. Landau R. Pharmacogenetics: implications for obstetric anesthesia. *Int J Obstet Anesth.* 2005 Oct;14(4):316-23.
12. Kharasch ED, Thummel KE. Identification of cytochrome P450 2E1 as the predominant enzyme catalyzing human liver microsomal defluorination of sevoflurane, and methoxyflurane. *Anesthesiology.* 1993 Oct;79(4):795-807.
13. Gentz BA, Malan TP Jr. Renal toxicity with sevoflurane: a storm in a teacup? *Drugs.* 2001;61(15):2155-62.
14. Watanabe J, Hayashi S, Nakachi K, Imai K, Suda Y, Sekine T, Kawajiri K. PstI and RsaI RFLPs in complete linkage disequilibrium at the CYP2E gene. *Nucleic Acids Res.* 1990 Dec 11;18(23):7194.
15. Hu Y, Oscarson M, Johansson I, Yue QY, Dahl ML, Tabone M, Arincò S, Albano E, Ingelman-Sundberg M. Genetic polymorphism of human CYP2E1: characterization of two variant alleles. *Mol Pharmacol.* 1997 Mar;51(3):370-6.
16. Preissner S, Kroll K, Dunkel M, Goldsobel G, Kuzmann D, Senger S, Günther S, Winnenburg R, Schroeder M, Preissner R. SuperCYP: a comprehensive database on Cytochrome P450 enzymes including a tool for analysis of CYP-drug interactions. *Nucleic Acids Res.* 2010 Jan;38(Database issue):D237-43.
17. Kharasch ED, Hankins DC, Thummel KE. Human kidney methoxyflurane and sevoflurane metabolism. Intrarenal fluoride production as a possible mechanism of methoxyflurane nephrotoxicity. *Anesthesiology.* 1995 Mar;82(3):689-99.
18. Kaymak C, Karahalil B, Ozcan NN, Oztuna D. Association between GSTP1 gene polymorphism and serum alpha-GST concentrations undergoing sevoflurane anaesthesia. *Eur J Anaesthesiol.* 2008 Mar;25(3):193-9.
19. Vanlersberghe C, Camu F. Propofol. *Handb Exp Pharmacol.* 2008;(182):227-52.
20. Yamanaka H, Nakajima M, Katoh M, Hara Y, Tachibana O, Yamashita J, McLeod HL, Yokoi T. A novel polymorphism in the promoter region of human UGT1A9 gene (UGT1A9*22) and its effects on the transcriptional activity. *Pharmacogenetics.* 2004 May;14(5):329-32.
21. Loryan I, Lindqvist M, Johansson I, Hiratsuka M, van der Heiden I, van Schaik RH, Jakobsson J, Ingelman-Sundberg M. Influence of sex on propofol metabolism, a pilot study: implications for propofol anesthesia. *Eur J Clin Pharmacol.* 2012 Apr;68(4):397-406.
22. Guitton J, Buronfosse T, Desage M, Flinois JP, Perdrix JP, Brazier JL, Beaune P. Possible involvement of multiple human cytochrome P450 isoforms in the liver metabolism of propofol. *Br J Anaesth.* 1998 Jun;80(6):788-95.
23. Franks NP. Molecular targets underlying general anaesthesia. *Br J Pharmacol.* 2006 Jan;147 Suppl 1:S72-81.
24. Davies PA, Hanna MC, Hales TG, Kirknrs EF. Insensitivity to anaesthetic agents conferred by a class of GABA (A) receptor subunit. *Nature.* 1997 Feb 27;385(6619):820-3.
25. Bali M, Akabas MH. Defining the propofol binding site location on the GABA A receptor. *Mol Pharmacol.* 2004 Jan;65(1):68-76.

26. Krasowski MD, Koltchine VV, Rick CE, Ye Q, Finn SE, Harrison NL. Propofol and other intravenous anesthetics have sites of action on the gamma-aminobutyric acid type A receptor distinct from that for isoflurane. *Mol Pharmacol*. 1998 Mar;53(3):530-8.
27. Tsang SY, Ng SK, Xu Z, Xue H. The evolution of GABA A receptor - like genes. *Mol Biol Evol*. 2007 Feb;24(2):599-610.
28. Richardson JE, Garcia PS, O'Toole KK, Derry JM, Bell SV, Jenkins A. A conserved tyrosine in the beta2 subunit M4 segment is a determinant of gamma-aminobutyric acid type A receptor sensitivity to propofol. *Anesthesiology*. 2007 Sep;107(3):412-8.
29. Iohom G, Ni Chonghaile M, O'Brien JK, Cunningham AJ, Fitzgerald DF, Shields DC. An investigation of potential genetic determinants of propofol requirements and recovery from anaesthesia. *Eur J Anaesthesiol*. 2007 Nov;24(11):912-9.
30. Kalow W. Interethnic variation of drug metabolism. *Trends Pharmacol Sci*. 1991 Mar;12(3):102-7.
31. Searle R, Hopkins PM. Pharmacogenomic variability and anaesthesia. *Br J Anaesth*. 2009 Jul;103(1):14-25.
32. Lin M, Aquilante C, Johnson JA, Wu R. Sequencing drug response with HapMap. *Pharmacogenomics J*. 2005;5(3):149-56.
33. Ingelman-Sundberg M, Gomez A. The past, present and future of pharmacoeugenomics. *Pharmacogenomics*. 2010 May;11(5):625-7.
34. Kacevska M, Ivanov M, Ingelman-Sundberg M. Perspectives on epigenetics and its relevance to adverse drug reactions. *Clin Pharmacol Ther*. 2011 Jun;89(6):902-7.
35. Villeneuve L, Girard H, Fortier LC, Gagné JF, Guillemette C. Novel functional polymorphisms in the UGT1A7 and UGT1A9 glucuronidating enzymes in Caucasian and African-American subjects and their impact on the metabolism of 7-ethyl-10-hydroxycamptothecin and flavopiridol anticancer drugs. *J Pharmacol Exp Ther*. 2003 Oct;307(1):117-28.
36. Saeki M, Saito Y, Jinno H, Sai K, Komamura K, Ueno K, Kamakura S, Kitakaze M, Shirao K, Minami H, Ohtsu A, Yoshida T, Saijo N, Ozawa S, Sawada J. Three novel single nucleotide polymorphisms in UGT1A9. *Drug Metab Pharmacokinet*. 2003;18(2):146-9.
37. Takahashi H, Maruo Y, Mori A, Iwai M, Sato H, Takeuchi Y. Effect of D256N and Y483D on propofol glucuronidation by human uridine 5'-diphosphate glucuronosyltransferase (UGT1A9). *Basic Clin Pharmacol Toxicol*. 2008 Aug;103(2):131-6.
38. Girard H, Villeneuve L, Court MH, Fortier LC, Caron P, Hao Q, von Moltke LL, Greenblatt DJ, Guillemette C. The novel UGT1A9 intronic I399 polymorphism appears as a predictor of 7-ethyl-10-hydroxycamptothecin glucuronidation levels in the liver. *Drug Metab Dispos*. 2006 Jul;34(7):1220-8.
39. Kansaku F, Kumai T, Sasaki K, Yokozuka M, Shimizu M, Tateda T, Murayama N, Kobayashi S, Yamazaki H. Individual differences in pharmacokinetics and pharmacodynamics of anesthetic agent propofol with regard to CYP2B6 and UGT1A9 genotype and patient age. *Drug Metab Pharmacokinet*. 2011;26(5):532-7.
40. Kirchheiner J, Klein C, Meineke I, Sasse J, Zanger UM, Mürdter TE, Roots I, Brockmüller J. Bupropion and 4-OH-bupropion pharmacokinetics in relation to genetic polymorphisms in CYP2B6. *Pharmacogenetics*. 2003 Oct;13(10):619-26.
41. Lang T, Klein K, Fischer J. Extensive genetic polymorphism in the human CYP2B6 gene with impact on expression and function in human liver. *Pharmacogenetics*. 2001 Jul;11(5):399-415.
42. Raftogianis RB, Wood TC, Otterness DM, Van Loon JA, Weinshilboum RM. Phenol sulfotransferase pharmacogenetics in humans: association of common SULT1A1 alleles with TS PST phenotype. *Biochem Biophys Res Commun*. 1997 Oct 9;239(1):298-304.
43. Traver RD, Siegel D, Beall HD, Phillips RM, Gibson NW, Franklin WA, Ross D. Characterization of a polymorphism in NAD(P)H: quinone oxidoreductase (DT-diaphorase). *Br J Cancer*. 1997;75(1):69-75.